



Hydraulic and electric control of cell spheroids

Charlie Duclut^a, Jacques Prost^{b,c}, and Frank Jülicher^{a,d,e,1}

^aMax-Planck-Institut für Physik Komplexer Systeme, 01187 Dresden, Germany; ^bLaboratoire Physico Chimie Curie, UMR 168, Institut Curie, PSL Research University, CNRS, Sorbonne Université, 75005 Paris, France; ^cMechanobiology Institute, National University of Singapore, 117411 Singapore, Singapore; ^dCenter for Systems Biology Dresden, 01307 Dresden, Germany; and ^eCluster of Excellence Physics of Life, Technische Universität Dresden, 01062 Dresden, Germany

Edited by David A. Weitz, Harvard University, Cambridge, MA, and approved March 28, 2021 (received for review November 10, 2020)

We use a theoretical approach to examine the effect of a radial fluid flow or electric current on the growth and homeostasis of a cell spheroid. Such conditions may be generated by a drain of micrometric diameter. To perform this analysis, we describe the tissue as a continuum. We include active mechanical, electric, and hydraulic components in the tissue material properties. We consider a spherical geometry and study the effect of the drain on the dynamics of the cell aggregate. We show that a steady fluid flow or electric current imposed by the drain could be able to significantly change the spheroid long-time state. In particular, our work suggests that a growing spheroid can systematically be driven to a shrinking state if an appropriate external field is applied. Order-of-magnitude estimates suggest that such fields are of the order of the indigenous ones. Similarities and differences with the case of tumors and embryo development are briefly discussed.

tissue biophysics | electrohydraulics | continuum theory of tissues | multicellular spheroids | tissue growth

Understanding how cells collectively organize to form complex structures and organs is the fundamental question raised in morphogenesis. This self-organization stems from the interplay of biochemical (1, 2) and mechanical (2–4), but also hydraulic (5, 6) and electrical, processes (7, 8). A long-standing paradigm in developmental biology is that cell chemical signals, in the form of morphogens, control cell growth and differentiation leading to tissue patterning (1). Although this biochemical signaling is of paramount importance in developmental control, it is now well established that mechanical forces between cells or mediated by the extracellular matrix can also provide regulatory cues that are equally important (3).

The crucial importance of hydraulics in morphogenesis, which should not come as a surprise given the large water content in tissues, has also been highlighted in multiple experiments. Hydraulic oscillations have for instance been shown to provide a robust mechanism for size control of the mouse embryo (9) and during the *Hydra* regeneration (10). The role of electrical signals in tissue patterning, although already studied by Roux (11) at the end of the 19th century, has gained a new interest only recently (7, 12, 13). In addition to its key function in receiving and relaying sensory information in the nervous system (14), bioelectricity has been shown to have a dramatic importance in large-scale patterning: An alteration of the electrical signaling in *Planaria* regeneration causes for instance the emergence of animals with multiple heads (7). Similarly, it has recently been observed that an external electric field can be used to reverse the morphogenetic fate of *Hydra* (15).

In the quest to understand how these different mechanisms come together to shape tissues and organs, simple cell aggregates such as spheroids have offered an appealing territory to observe tissue development and to formulate hypotheses on the underlying mechanisms. Remarkably, a rich behavior is observed even in single-cell type spheroids, as in for instance their ability to pump fluid and form liquid-filled lumens (16, 17). More recently, organoids have received increasing attention as they can recapitulate complex mor-

phogenetic processes in a relatively simple and controllable environment (18, 19).

To unravel the connections between biochemical signals and tissue mechanics, mechanical perturbations of organoids and cell spheroids can be performed. For instance, atomic force microscopy has been used to probe the mechanical properties of mammary organoids (20) and revealed the importance of both extracellular matrix stiffness and laminin signaling to maintain tissue integrity. Perturbation of the osmotic pressure around cell spheroids using large molecules of dextran has also highlighted the importance of isotropic stress in tissue growth (21, 22).

A broad understanding of tissue mechanics therefore requires us to consider tissue electrohydraulic properties and to be able to perturb tissues by electric or hydraulic means. In this theoretical work, we study the response of a cell spheroid to electric and hydraulic perturbations. We propose an experimental setup where a pipette or drain is used to impose a fluid flow or an external electric current through a micrometric drain. Our work shows that one could control the size of a cellular assembly using such a setup.

Our theoretical approach relies on generic features of the physics of the electro-hydrodynamic phenomena at play within tissues. We adopt a coarse-grained approach of tissues, which describes many cells and their microenvironments as a continuum with active material properties (23–25). Cell mechanical characteristics, fluid pumping, ion transport, and electrical properties are thus considered in a unified framework (26, 27) that we use to analyze spheroid response to an external perturbation.

In particular, we highlight in the following that steady external flow or electric current imposed by a drain can systematically drive a proliferating spheroid to degeneracy. Such a technique

Significance

In addition to generating forces and reacting to mechanical cues, tissues are capable of actively pumping fluid and creating electric current. In this work, we examine how a hydraulic or electrical perturbation, imposed, for instance, by a drain of micrometric diameter, can be used to perturb tissue growth. We address this issue in a continuum description of a spherical cell assembly that includes the mechanical, electrical, and hydraulic properties of the tissue. This approach allows us to discuss and quantify the effect of electrohydraulic perturbations on the long-time states of the tissue. We highlight that a moderate external pumping or electric current can drive a proliferating spheroid to decay. We propose that this could have applications in medicine.

Author contributions: C.D., J.P., and F.J. designed research, performed research, analyzed data, and wrote the paper.

The authors declare no competing interest.

This article is a PNAS Direct Submission.

Published under the PNAS license.

¹To whom correspondence may be addressed. Email: julicher@pks.mpg.de.

This article contains supporting information online at <https://www.pnas.org/lookup/suppl/doi:10.1073/pnas.2021972118/-DCSupplemental>.

Published May 4, 2021.

could be relevant in a medical context where it could be used to suppress cancerous tumors.

Methods for suppressing malignant tumors are numerous: A first path to control tumor size is to use drugs to disturb the chemical regulation of cancerous cells to prevent them from proliferating (chemotherapy). Immunotherapy has offered a strong alternative in mitigating cancer by stimulating the immune system to suppress tumors (28). In addition, radiotherapy—which allows the suppression of the tumor by damaging the genetic material of cancerous cells through radiations—has also proved effective for tumor removal (29).

Physics-based methods can also be used to provide new techniques for directly suppressing the cancerous tissues. High-intensity focused ultrasounds can for instance be used to locally overheat the cancerous cells (30). Recent experiments also suggest that lower-intensity ultrasound waves could be used to strain and suppress mechanically cancerous tissues (31, 32), and low-frequency ultrasounds could be used to increase selectivity (32). Electrical perturbations have also been used: Cancerous tissues can for instance be successfully suppressed using irreversible electroporation (33) by applying large voltage pulses in the tissue. More recently, microelectrodes have been used in electrolytic ablation methods to locally change pH and kill a cancerous mass (34).

Compared to other treatments and ablation techniques (35), direct electrohydraulic perturbations of cancerous tissues have, however, remained largely unexplored. With the theoretical study that we present here, we aim at highlighting potential methods for controlling the size of cell spheroids that could also be used to suppress cancerous tumors.

Continuum Model of a Spheroid with a Drain

Following refs. 26 and 27, we consider the tissue at a coarse-grained level, such that individual cells are not described but the tissue as a whole is studied as a continuum material with active electrical, hydraulic, and mechanical properties. To capture the hydraulic properties of the tissue, we also adopt a two-fluid description, where the cells, which form the first fluid, are permeated by the interstitial fluid (36). In the long-time limit (several days or weeks) that we consider here, cells are able to reorganize and to relax the internal stresses within the tissue, such that an effective description of the tissue as a viscous active fluid at long times is used (25).

To analyze specifically the effect of a drain on a cell spheroid, we consider a spherical tissue of radius R_2 enclosing a spherical lumen of radius R_1 (Fig. 1). A drain of inner radius R_d is inserted

inside the spheroid and can be used to impose an external flow or an external current. We consider, despite the presence of the drain, a system with spherical symmetry. The description we propose in the following is therefore effectively one dimensional and depends only on the distance r to the spheroid center.

Directional ion pumping through the spheroid is achieved if cells have a polarity. We thus define a cell polarity field \mathbf{p} with unit norm that we assume, for simplicity, to be oriented along the radial direction: $\mathbf{p} = \mathbf{e}_r$. Cells also display a nematic ordering—due for instance to an anisotropy in their shape—that we describe with the nematic tensor $q_{\alpha\beta}$ (Greek indexes indicate Cartesian coordinates). We assume in the following that the nematic ordering is defined by the same preferred axis as the polarity of the cells: $q_{\alpha\beta} = p_\alpha p_\beta - (1/3)\delta_{\alpha\beta}$. This situation is for instance observed in colon carcinoma cell spheroids (37). Finally, the consideration of ion pumping within the tissue requires the introduction of an electric field \mathbf{E} and an electric current density \mathbf{j} , which obey a generalized Ohm law as we discuss in the following section.

Tissue Mechanical, Hydraulic, and Electrical Properties. The total tissue stress is decomposed as $\sigma_{\alpha\beta} = \sigma_{\alpha\beta}^c + \sigma_{\alpha\beta}^f$, where here and in the following the superscripts c, f stand for the cells and interstitial fluid contributions, respectively. Neglecting inertia and in the absence of external bulk forces, force balance within the tissue reads

$$\partial_\beta \sigma_{\alpha\beta}^c + f_\alpha = 0, \quad [1a]$$

$$\partial_\beta \sigma_{\alpha\beta}^f - f_\alpha = 0, \quad [1b]$$

where f_α represents internal forces between the cells and the interstitial fluid, and where summation over repeated indexes is implied.

To obtain the dynamics of the spheroid, that is, the time evolution of the inner and outer radii R_1 and R_2 , we now need to specify the material properties (or constitutive equations) of the tissue. In our coarse-grained description, the interstitial fluid is driven by pressure gradients, while fluid viscosity contributes to the internal forces f_α between fluid and cells. We therefore write the fluid stress as $\sigma_{\alpha\beta}^f = -P^f \delta_{\alpha\beta}$. For the cell stress, the description of the material properties is made easier by decomposing the stress into a traceless symmetric part $\tilde{\sigma}_{\alpha\beta}^c$ and an isotropic part $\sigma^c \delta_{\alpha\beta}$. The constitutive equation for the isotropic part then reads (26, 27)

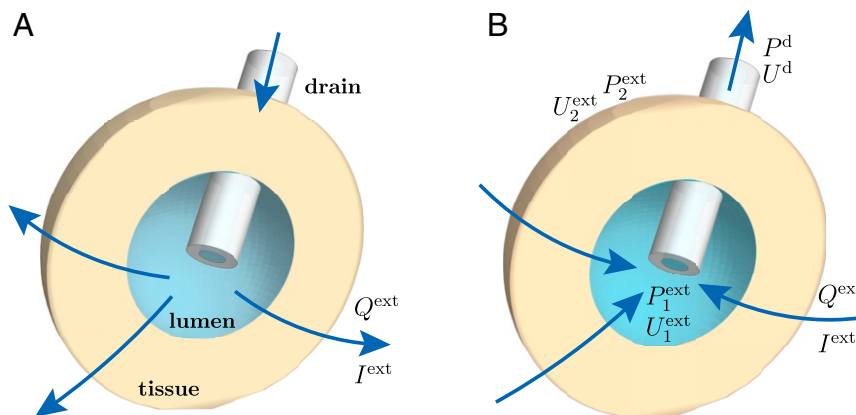


Fig. 1. Sketch of a spheroid with drain. The drain is used to impose an electric current I^{ext} or a volumetric flow rate Q^{ext} , driving a fluid flow or current density throughout the cell spheroid. (A) Flows from inside of the spheroid to outside correspond to $Q^{\text{ext}}, I^{\text{ext}} > 0$. (B) Flows from outside to inside correspond to $Q^{\text{ext}}, I^{\text{ext}} < 0$. The pressures inside the lumen, outside the spheroid, and at the outer end of the drain are denoted $P_1^{\text{ext}}, P_2^{\text{ext}}$, and P^d , respectively, and correspondingly for the electric potential: $U_1^{\text{ext}}, U_2^{\text{ext}}$, and U^d .

$$\sigma^c + P_h^c = \bar{\eta} v_{\gamma\gamma}^c - \nu_0 \bar{\sigma}_{\alpha\beta}^c q_{\alpha\beta} - \nu_1 p_\alpha E_\alpha - \nu_2 p_\alpha V_\alpha, \quad [2a]$$

where $-\sigma^c$ is the local pressure and we have introduced the cell strain-rate tensor $v_{\alpha\beta}^c = (\partial_\alpha v_\beta^c + \partial_\beta v_\alpha^c)/2$ and where we have defined $V_\alpha = v_\alpha^c - v_\alpha^f$. The bulk viscosity $\bar{\eta}$ is a familiar term that is also present for a passive fluid, except that its microscopic origin is different for a tissue. The homeostatic pressure P_h^c is a property specific for tissues that results from a balance of cell growth and cell death (38). In addition to these terms, we have included additional terms in the isotropic stress: Cell anisotropies can couple to the anisotropic stress, resulting in the term proportional to ν_0 . The terms proportional to ν_1 and ν_2 represent bioelectric and biohydraulic stresses induced by a coupling to the electric field or by a (relative) fluid flow, respectively. A similar expansion for the traceless anisotropic part of the stress tensor reads

$$\tilde{\sigma}_{\alpha\beta}^c = 2\eta \tilde{v}_{\alpha\beta}^c + \zeta q_{\alpha\beta} - \nu_3 [E_\alpha p_\beta]_{st} - \nu_4 [V_\alpha p_\beta]_{st}, \quad [2b]$$

where $\tilde{v}_{\alpha\beta}^c$ is the traceless part of the cell strain-rate tensor, and we have moreover defined the symmetric traceless part of a dyadic product of vectors: $[A_\alpha B_\beta]_{st} \equiv A_\alpha B_\beta + A_\beta B_\alpha - (2/3) A_\gamma B_\gamma \delta_{\alpha\beta}$. The coefficient η is the usual shear viscosity while the other terms represent active couplings, and the terms proportional to ν_3 and ν_4 are the anisotropic counterpart of the terms proportional to ν_1 and ν_2 in Eq. 2a. The active stress $\zeta q_{\alpha\beta}$ is a hallmark of active systems and shows the ability of cells to generate anisotropic stresses due to cell division or contraction of their cytoskeleton (39, 40). This active stress can be regulated by the cells and we therefore consider that it depends on the local pressure at linear order as

$$\zeta = \zeta_0 - \zeta_1 (\sigma^c + P_h^c). \quad [2c]$$

We emphasize that the cell stress constitutive equations (Eqs. 2a–2c) reflect the effective viscous properties of tissues at long time as a consequence of cellular growth and death. We review the derivation of these constitutive equations in *SI Appendix*. A key feature of our work is that flows and electric fields do influence cell division and death and therefore growth and shrinkage of the spheroid (26, 27, 41).

The constitutive equation for the internal force density between cells and interstitial fluid including all of the linear terms allowed by symmetry can be written as*

$$f_\alpha = -\kappa (v_\alpha^c - v_\alpha^f) + \lambda_1 p_\alpha + \lambda_2 E_\alpha + \lambda_3 \partial_\beta q_{\alpha\beta}. \quad [3]$$

The first term $-\kappa (v_\alpha^c - v_\alpha^f)$ accounts for the friction between the interstitial fluid and the cells and leads to Darcy's law (42) in the description of porous materials. The permeation coefficient can be estimated as $\kappa \simeq \eta^f / a^2$, where η^f is the interstitial fluid viscosity and a a typical interstitial distance. The term $\lambda_1 p_\alpha$ accounts for active fluid pumping by the cells, while the terms proportional to λ_2 correspond to the electroosmotic contribution. The last term, proportional to λ_3 , characterizes a differential pumping term due to the bending of the cells (43).

To complete the tissue properties description, one finally needs to specify the constitutive equation for the electric current density*

$$j_\alpha = -\bar{\kappa} (v_\alpha^c - v_\alpha^f) + \Lambda_1 p_\alpha + \Lambda_2 E_\alpha + \Lambda_3 \partial_\beta q_{\alpha\beta}, \quad [4]$$

where Λ_2 is the electric conductivity of the tissue. The term proportional to $\bar{\kappa}$ characterizes the current due to the (rela-

* Note that additional terms must be added to Eqs. 3 and 4 for a system lacking spherical symmetry or for a system where the polarity is not purely radial with unit norm; see *Appendix A* for details.

tive) flow of ions between cells as a consequence of a reverse electroosmotic effect (44). The coefficient Λ_1 characterizes the contribution of ion pumping to the electric current, while the coefficient Λ_3 is an active flexoelectric coefficient. It indicates that a spatially nonuniform cell polarity orientation is obtained in response to an electric field and has been shown to play a crucial role in the nucleation of a lumen in spherical cell aggregates (27).

Drain Description. The drain can be used to impose an external fluid flow or an external electric current in two equivalent ways: either by imposing directly a volumetric flow rate Q^{ext} and electric current I^{ext} through the drain or, alternatively, by applying a pressure difference $\Delta P = P^d - P_2^{\text{ext}}$ (with P_2^{ext} the pressure at the outer spheroid boundary and P^d the pressure at the outer end of the drain; Fig. 1) and an electric potential difference $\Delta U = U^d - U_2^{\text{ext}}$ (with U_2^{ext} the electric potential at the outer spheroid boundary and U^d the electric potential at the outer end of the drain).

In both cases, the imposed external fluid flow and external electric current density at the lumen boundary read

$$Q^{\text{ext}} = 4\pi R_1^2 v_1^{\text{ext}}, \quad \text{and} \quad I^{\text{ext}} = 4\pi R_1^2 j_1^{\text{ext}}, \quad [5]$$

where v_1^{ext} and j_1^{ext} are the fluid velocity and electric current density at the boundary between the spheroid and the lumen.

In the following, we focus on the case where Q^{ext} and I^{ext} are imposed externally. The case of a pressure difference or electric potential difference imposed by the drain is discussed in *Appendix C*.

Continuity Equations and Boundary Conditions. If cell density and interstitial fluid density are equal and constant, which we assume in the following, then the total volume flux $v_\alpha = \phi v_\alpha^c + (1 - \phi) v_\alpha^f$ is divergence-free: $\partial_\alpha v_\alpha = 0$ (see *SI Appendix* for details). In the presence of the drain, which imposes a nonvanishing fluid velocity at the inner boundary of the spheroid, the integration of the total flow incompressibility in spherical coordinates yields a relation between the fluid velocity v_r^f and cell velocity v_r^c in the tissue, which reads

$$v_r^f = \frac{v_1^{\text{ext}}}{1 - \phi} \left(\frac{R_1}{r} \right)^2 - \frac{\phi}{1 - \phi} v_r^c. \quad [6]$$

Similarly, charge conservation in the quasistatic limit $\partial_\alpha j_\alpha = 0$ can be integrated in the case where an external current density j_1^{ext} is imposed on the inner boundary, yielding the current density $j_r = j_1^{\text{ext}} (R_1/r)^2$ throughout the tissue.

The spheroid is surrounded by an external fluid both inside (in the lumen) and outside. This fluid exerts a hydrostatic pressure on the tissue that is balanced by the tissue surface tension and by the total normal stress at the boundaries:

$$-\sigma_{rr}^c(R_1) + P^f(R_1) = P_1^{\text{ext}} - 2\gamma_1/R_1, \quad [7a]$$

$$-\sigma_{rr}^c(R_2) + P^f(R_2) = P_2^{\text{ext}} + 2\gamma_2/R_2, \quad [7b]$$

where we have introduced the inner and outer tissue surface tensions γ_1 and γ_2 . Fluid exchange between the spheroid and the outside is driven by osmotic conditions:

$$v_1^{\text{ext}} - dR_1/dt = +K_1 \left[(P_1^{\text{ext}} - P^f(R_1)) - \Pi_1^{\text{ext}} \right] + J_{p,1}, \quad [8a]$$

$$v_2^{\text{ext}} - dR_2/dt = -K_2 \left[(P_2^{\text{ext}} - P^f(R_2)) - \Pi_2^{\text{ext}} \right] - J_{p,2}. \quad [8b]$$

Here, $K_{1,2}$ are the permeabilities of the interfaces to water flow. The fluxes $J_{p,1}$ and $J_{p,2}$ can be nonzero as a result of

active pumps and transporters that maintain an osmotic pressure difference and act effectively as water pumps. We have also introduced v_2^{ext} , the external flow imposed at outer spheroid boundary. Conservation of the volumetric flow directly yields $v_2^{\text{ext}} = v_1^{\text{ext}}(R_1/R_2)^2$.

The normal velocity of the cells at the boundaries has to match the growth of the spheroid radii. An increased cell proliferation in a thin surface layer has been observed in growing spheroids (21, 22, 45). We thus allow for a thin surface layer of cells, both facing outside and to the lumen, to have a growth rate that differs from the bulk. The cell velocity boundary conditions then read

$$v_r^c(R_1) = dR_1/dt + v_1, \quad [9a]$$

$$v_r^c(R_2) = dR_2/dt - v_2, \quad [9b]$$

where $v_i = \delta k_i n_i^c h / n^c$ with h the thickness of the boundary layers, and n_i^c and δk_i are the cell number density and the cell growth rate in the surface layers, respectively.

Dynamics of the Spheroid Growth and Orders of Magnitude

We have introduced in the previous section a model for a spherical spheroid with a drain. Solving the force balance Eq. 1 together with the boundary conditions Eqs. 7–9 then allows us to obtain the dynamics of the inner $R_1(t)$ and outer $R_2(t)$ radii of the spheroid in the quasistatic limit. We obtain two coupled nonlinear differential equations for the spheroid radii, Eqs. 23a and 23b (Appendix B), which have been studied in ref. 27 in the absence of a drain. As we will see in the following, imposing an external fluid flow or electric current has dramatic consequences for the spheroid growth and can be used to control its size.

Before discussing how the presence of external fields modifies the dynamics of a spheroid and which protocols can be used to control its growth, we first use our model to discuss the orders of magnitude of the external flux and electric current for which we expect a significant change in the spheroid dynamics. To obtain these estimates, we assume that the lumen size is small and use $R_1 = 0$ in Eq. 23b that describes the dynamics of the spheroid. We then compare in this equation the stresses generated by imposed flows or electric currents to the stresses stemming from internal activity in the absence of flows and currents. We find that the external volumetric flow required to significantly perturb the spheroid is of the order

$$Q^{\text{ext}} \simeq \sigma^{\text{typ}} \frac{4\pi(1-\phi)}{\kappa^{\text{eff}}} R_2, \quad [10]$$

where $\kappa^{\text{eff}} = \kappa - \bar{\kappa}\lambda_2/\Lambda_2$ is an effective permeation coefficient and σ^{typ} is a typical scale of tissue stress in the absence of flows and currents. Note that the appearance of κ^{eff} in Eq. 10 indicates that the finite bulk permeability of the spheroid governs the effects of the external flow imposed via the drain. Note, however, that for smaller spheroids of size $R_2 \lesssim (1-\phi)/\kappa^{\text{eff}} K_2 \simeq 10 - 50 \mu\text{m}$, the flow-induced effects are not dominated by bulk permeation but rather by surface permeation and we have $Q^{\text{ext}} \simeq 4\pi K_2 \sigma^{\text{typ}} R_2^2$. We can estimate the external electric current required to perturb the spheroid:

$$I^{\text{ext}} \simeq 4\pi \sigma^{\text{typ}} \Lambda_2 R_2 / \lambda_2. \quad [11]$$

One can use experimental values and order-of-magnitude estimates of the parameters that appear in Eqs. 10 and 11 (Table 1). We estimate that a volumetric flow $Q^{\text{ext}} \simeq 10^3 - 10^5 \mu\text{m}^3/\text{s}$ is sufficient to observe a significant change in the dynamics of a millimeter-sized spheroid, and this volumetric flow scales linearly with the size of the spheroid. Similarly, electric current of the order $I^{\text{ext}} \simeq 1 - 100 \text{ nA}$ can perturb the dynamics of a millimeter-sized spheroid (see Fig. 6 in Appendix E).

Table 1. Experimental values and references (left columns) and estimated values (right columns) of the phenomenological parameters of the model appearing in the constitutive equations

Parameters	Experimental values	Parameters	Estimations (26, 27)
η (51)	$10^4 \text{ Pa}\cdot\text{s}$	$\bar{\kappa}$	$10^3 \text{ A}\cdot\text{s}/\text{m}^3$
$\bar{\eta}$ (21)	$10^9 \text{ Pa}\cdot\text{s}$	λ_1	$-10^8 \text{ N}/\text{m}^3$
$\gamma_{1,2}$ (51)	$10^{-3} \text{ N}/\text{m}$	λ_2	$-10^6 \text{ N}\cdot\text{m}^{-2}\cdot\text{V}^{-1}$
κ^{-1} (52)	$10^{-13} \text{ m}^2\cdot\text{Pa}^{-1}\cdot\text{s}^{-1}$	λ_3	$-10^3 \text{ N}/\text{m}^2$
$\Pi_{1,2}^{\text{ext}}$ (53)	10^3 Pa	Λ_1	$1 \text{ A}/\text{m}^2$
$v_{1,2}$ (45)	$10^{-10} \text{ m}/\text{s}$	Λ_2	$10^{-2} \text{ A}\cdot\text{V}^{-1}\cdot\text{m}^{-1}$
P_h^c (21)	-10^3 Pa	Λ_3	$10^{-5} \text{ A}/\text{m}$
ζ_0 (45)	10^3 Pa	ν_1	$10^1 \text{ N}\cdot\text{m}^{-1}\cdot\text{V}^{-1}$
ζ_1 (45)	-10^{-1}	ν_2	$10^8 \text{ Pa}\cdot\text{s}/\text{m}$
R_d	$5 \cdot 10^{-6} \text{ m}$	ν_3	$1 \text{ N}\cdot\text{m}^{-1}\cdot\text{V}^{-1}$
		ν_4	$10^7 \text{ Pa}\cdot\text{s}/\text{m}$
		$K_{1,2}$	$10^{-10} \text{ m}\cdot\text{Pa}^{-1}\cdot\text{s}^{-1}$
		$J_{p,1,2}$	$10^{-11} \text{ m}/\text{s}$
		$1 - \phi$	10^{-1}
		ν_0	1

Externally imposed flows and electric currents could therefore be used to induce a change in the spheroid behavior. In the following sections, we focus in more detail on how these external perturbations can be used to control the size of the spheroid.

Hydraulic and Electric Control of the Size of a Spheroid

Examples of Protocols for Spheroid Suppression. Using our model and solving numerically Eqs. 23a and 23b (Appendix D), we can analyze various protocols for the suppression of a spheroid. The dynamics of the spheroid and its lumen, $R_2(t)$ and $R_1(t)$, are therefore studied for different external flow protocols $Q^{\text{ext}}(t)$ and electric current protocols $I^{\text{ext}}(t)$. To keep the discussion as general as possible, we introduce dimensionless quantities: a dimensionless radius $r_2(\hat{t}) = R_2(t)/R_0$, time $\hat{t} = t/\tau_0$, external volumetric flow $\hat{Q} = Q^{\text{ext}}/Q_0$, and electric current $\hat{I} = I^{\text{ext}}/I_0$, where we have defined

$$R_0 = K_2 \bar{\eta}, \quad \tau_0 = \bar{\eta} / |P_2^{\text{eff}}|, \quad [12]$$

$$Q_0 = 4\pi \bar{\eta}^2 K_2^3 |P_2^{\text{eff}}|, \quad I_0 = 4\pi \Lambda_2 \bar{\eta} K_2 |P_2^{\text{eff}}| / |\lambda_2|.$$

Note that the effective pressure P_2^{eff} introduced above is a modification of the homeostatic pressure P_h^c by the external osmotic pressure and by electric and active contributions. Its expression can be found in Table 2 and in Appendix B.

We display in Fig. 2 different protocols that can be applied to a growing spheroid to suppress it. If the external flow magnitude is sufficiently large and if it is applied long enough, the interventions are successful and lead to the spheroid suppression. Fig. 2B shows an example of a successful suppression of the spheroid as a sufficiently strong flow has been applied until the spheroid is suppressed. A larger flow magnitude leads to a faster spheroid suppression (Fig. 2C). We also show in Fig. 2D an example of a successful suppression protocol for which the magnitude of the flow is lowered as the spheroid size decreases.

Conversely, if the magnitude of the imposed external flow is not sufficient, or if the duration of the flow is too short, the protocol can be unsuccessful in suppressing the spheroid. The spheroid may only have a slower growth rate (Fig. 2E) or shrink significantly but resume growing as soon as the external flow is turned off (Fig. 2F). Note that for any spheroid size, there exists a critical flow magnitude $|Q_c|$ such that imposing a steady flow at magnitude $|Q^{\text{ext}}| > |Q_c|$ will eventually lead to the spheroid suppression, while $|Q^{\text{ext}}| < |Q_c|$ will be unsuccessful. Fig. 2B shows a protocol with $|Q^{\text{ext}}| \simeq |Q_c|$.

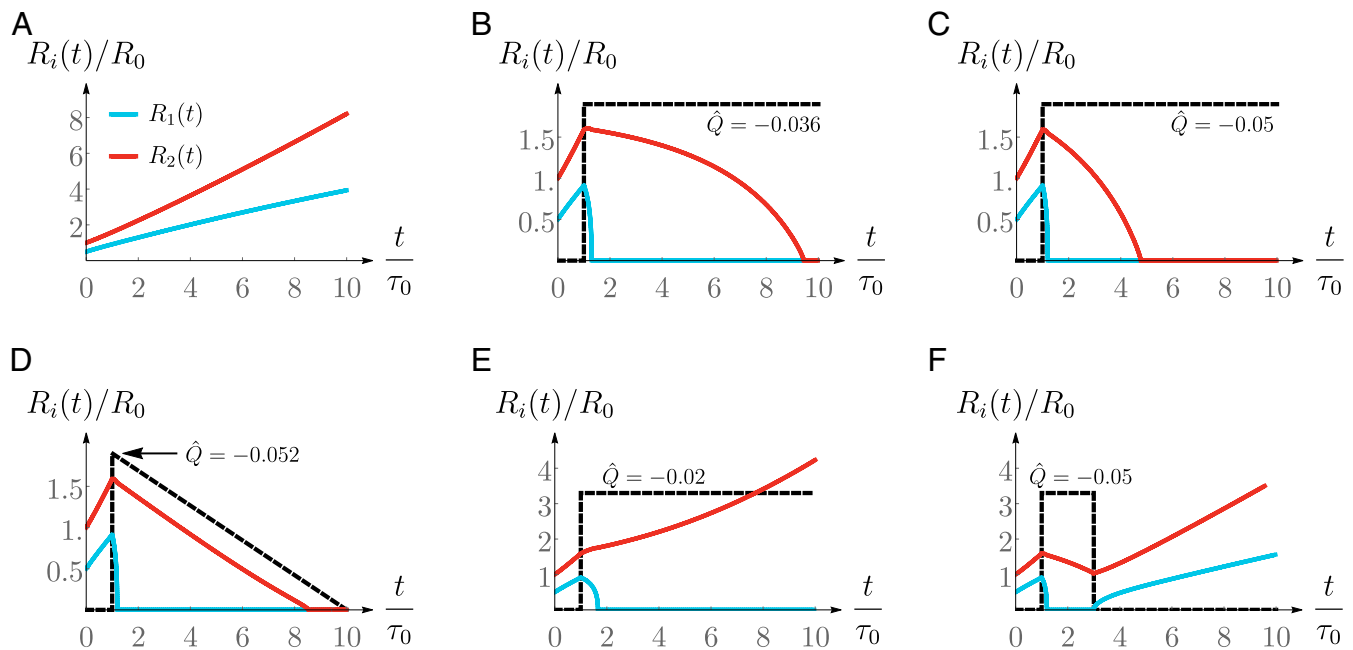


Fig. 2. Spheroid size R_2 (red curve) and lumen size R_1 (blue curve) normalized to a characteristic size R_0 as a function of the normalized time $\hat{t} = t/\tau_0$ for different external flow protocols $\hat{Q}(\hat{t}) = Q^{\text{ext}}(t)/Q_0$ (dashed black curve; its magnitude is not at scale). The definitions of τ_0 , R_0 , and Q_0 are given in Eq. 12. (A) Spheroid growth without external intervention. (B–D) Successful protocols: The spheroid is suppressed. (E and F) Unsuccessful protocols: The spheroid continues to grow. These plots are obtained by solving the dynamics equations Eqs. 23a and 23b with the parameter values given in Table 3.

In the examples above we have focused on the case where an external volumetric flow is imposed. The same analysis and similar procedures can be used in the case where an external current $I^{\text{ext}}(t)$ is imposed (Fig. 3). We show in Fig. 3B a protocol that leads to the suppression of the spheroid. Note that an increased shrinking is obtained if one applies simultaneously an electric current and an external flow, as both effects are additive. Such intervention is displayed in Fig. 3C where we observe a faster shrinking due to external flow in addition to the application of an electric current.

Importantly, we emphasize that the protocols we are discussing here are slow and take place on long time scales: Using parameter values displayed in Table 3, the examples shown in Figs. 2 and 3 correspond to $R_0 \simeq 1$ cm and $\tau_0 \simeq 10$ days. This shows that the suppression of the spheroid requires a slow and steady flow or the application of a small electric current for several weeks.

Spheroid Dynamics with External Flows and Currents. We now discuss quantitatively how a drain can be used to control the size

of a spheroid. To keep the discussion as simple as possible, we consider here the limit of a spheroid enclosing a small lumen (or lumenless). In the limit of a small lumen compared to spheroid size $R_1 \ll R_2$, the differential equation that describes the dynamics of the spheroid shrinks to (Appendix B)

$$\frac{dr_2}{d\hat{t}} = \frac{w_0 + (\hat{v}_2 + w_1/2)r_2 - \delta_2 r_2^2 + \hat{\lambda} r_2^3/4}{r_2(3 + r_2)}, \quad [13]$$

where we use the dimensionless radius r_2 and time \hat{t} introduced in the previous section. The dimensionless parameters $w_{0,1}$ describe the effects of an externally imposed flow or electric current. If they are set to zero, Eq. 13 reduces to the dynamics of a lumenless spheroid without drain as studied in ref. 27. They are defined as $w_n = j_n \hat{I} + u_n \hat{Q}$ with $n = 0, 1$ and where \hat{Q} and \hat{I} are the dimensionless volumetric flow and electric current, respectively, as defined above. The effects of an imposed external flow are captured by the coefficients

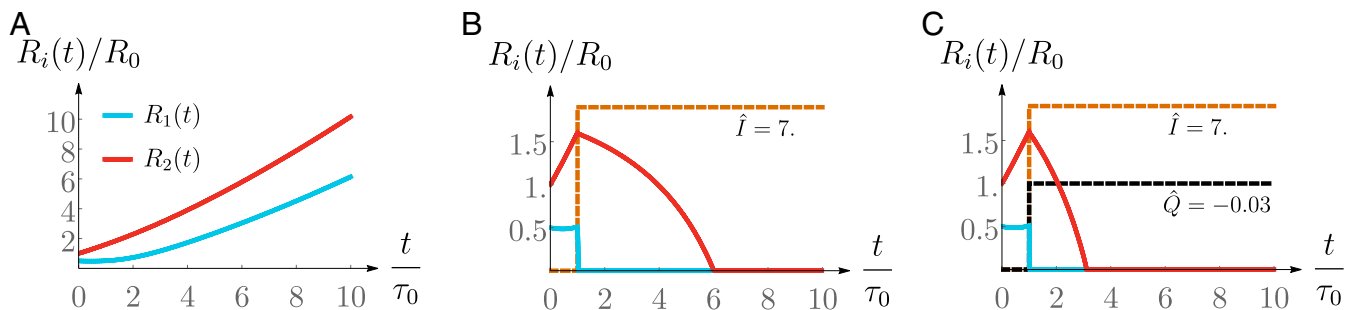


Fig. 3. Dimensionless spheroid size R_2/R_0 (red curve) and lumen size R_1/R_0 (blue curve) as a function of dimensionless time $\hat{t} = t/\tau_0$ in the presence of an imposed electric current $\hat{I}(\hat{t}) = I^{\text{ext}}(t)/I_0$ and imposed flow $\hat{Q}(\hat{t}) = Q^{\text{ext}}(t)/Q_0$ (dashed brown and black curves; their magnitude is not at scale). The definitions of τ_0 , R_0 , Q_0 , and I_0 are given in Eq. 12. (A) Spheroid growth without external intervention. (B) Successful protocol with external electric field. (C) Successful protocol with external electric field and flow. Parameter values used for these plots can be found in Table 3.

$$u_1 = \frac{\kappa^{\text{eff}} \bar{\eta} K_2^2}{1 - \phi}, \quad u_0 = 1 - \beta_u. \quad [14]$$

Here, u_1 describes a permeation effect due to the finite permeability of the spheroid to fluid flows and involves the ratio of the surface permeability K_2 and of the (effective) bulk permeability $\propto 1/\kappa^{\text{eff}}$. The parameter u_0 includes direct effects of the external flow—proportional to Q^{ext}/K_2 in the dimensional equation corresponding to $u_0 = 1$ —which reflects the spheroid volume change imposed by the external flow due to tissue incompressibility (Appendix B). We have furthermore defined

$$\beta_u = \frac{K_2}{1 - \phi} \left(3\nu_2 - 4\nu_0\nu_4 + \frac{\bar{\kappa}}{\Lambda_2} (3\nu_1 - 4\nu_0\nu_3) \right), \quad [15]$$

which represents the bioelectric and biohydraulic contribution. Indeed, this term is a sum of terms proportional to the parameters ν_i and thus stems from the coupling between the electric field (or the interstitial fluid flow) and the cell polarity that appears in the cell stress.

Imposing an external electric current I^{ext} also contributes to the spheroid growth control via the parameters

$$j_1 = 1, \quad j_0 = \beta_j. \quad [16]$$

The coefficient $j_1 = 1$ corresponds to electroosmotic flow due to the imposed current. The coefficient j_0 accounts for bioelectric and biohydraulic contributions, where

$$\beta_j = \frac{3\nu_1 - 4\nu_0\nu_3}{\lambda_2 \bar{\eta} K_2}. \quad [17]$$

Note that since the coefficients $\beta_{u,j}$ are a sum of phenomenological parameters for which we have only order of magnitude estimates, it is difficult to obtain a reliable estimate of their magnitude and even of their sign. However, estimating upper bounds for $\beta_{u,j}$ suggests that these bioelectric and biohydraulic contributions are small compared to the other effects. A further discussion of these contributions requires precise estimates of the bioelectric and biohydraulic couplings that could be obtained from experiments on spheroids in the presence of a drain.

In Eq. 13, we have used the definitions (27)

$$\hat{\lambda} = \frac{\lambda^{\text{eff}} \bar{\eta} K_2}{|P_2^{\text{eff}}|}, \quad \delta_2 = \frac{P_2^{\text{eff}}}{|P_2^{\text{eff}}|}, \quad \hat{v}_2 = \frac{v_2^{\text{eff}}}{K_2 |P_2^{\text{eff}}|}, \quad [18]$$

$$v_2^{\text{eff}} = 3v_2 - \frac{2(\gamma_2^{\text{app}} + \gamma_0^{\text{app}})}{\bar{\eta}}.$$

Here, λ^{eff} is an effective pumping coefficient and γ_0^{app} are apparent tensions defined in Appendix B. The apparent surface tension γ_2^{app} is a modification of the tissue surface tension γ_2 stemming from the flexoelectric term proportional to Λ_3 . This flexoelectric contribution plays a crucial role in lumen nucleation (27). In the dynamics of the outer radius of the spheroid, its effect is, however, minor. Finally, the effective pumping λ^{eff} combines the active pumping λ_1 and an electric contribution $\Lambda_1 \lambda_2 / \Lambda_2$ due to electroosmosis. The parameters introduced above are summarized for convenience in Table 2, and their corresponding values can be computed using Table 1.

Control of the Spheroid Dynamics. We use Eq. 13 to discuss how external currents and flows can be used to control the growth and shrinkage of a spheroid. Note that whether the spheroid grows ($dr_2/dt > 0$) or shrinks ($dr_2/dt < 0$) depends only on the numerator of Eq. 13.

Fig. 4 shows phase-space trajectories of the outer radius r_2 for spheroids that are able to grow in the absence of a drain. Fig. 4A displays the case without external flow or current. The following scenarios are possible: The spheroid may be growing with an unstable fixed point at $r_2 = 0$ (green curve); alternatively, there might be an additional stable fixed point corresponding to a steady state, either with finite radius $r_2^* \neq 0$ (blue curve) or with vanishing radius $r_2^* = 0$ (orange curve). Imposing an external flow implies nonzero values of w_0 and w_1 , which are negative in the case where fluid flows out of the lumen (depicted in Fig. 1B). The case of a nonvanishing value of w_0 is displayed in Fig. 4B: In this case, the trajectories are shifted downward by an amount w_0 . In this case, even the growing spheroid (green curve) has a critical radius below which it shrinks (open green circle). The stable steady state (blue curve) has moved to $r_2^* = 0$. Fig. 4C shows the effect of a nonvanishing value of w_1 : The slope of the phase-space trajectories is modified. Note that this

Table 2. Summary of the effective parameters introduced in the text, their definition, and the corresponding equation in the text

Parameter	Definition	Equation	Description
κ^{eff}	$\kappa - \bar{\kappa} \lambda_2 / \Lambda_2$	Below Eq. 10	Effective permeation coefficient
$P_{1,2}^{\text{eff}}$	$\Pi_{1,2}^{\text{ext}} - P_h^c - \frac{f_{p,1,2}}{\kappa_{1,2}} - \frac{2}{3} \left(\zeta_0 \nu_0 + \lambda_3 + \frac{\Lambda_3 \lambda_2 + (3\nu_1/2 - 2\nu_0\nu_3)\Lambda_1}{\Lambda_2} \right)$	Eq. 25a	Effective pressure
$\gamma_{1,2}^{\text{app}}$	$\gamma_{1,2} \mp 4\nu_3 \Lambda_3 / \Lambda_2$	Eq. 25b	Apparent surface tension
γ_0^{app}	$(3\nu_1/2 - 2(2 + \nu_0)\nu_3)(\Lambda_3 / \Lambda_2)$	Eq. 25b	Apparent surface tension
λ^{eff}	$\lambda_1 - \Lambda_1 \lambda_2 / \Lambda_2$	Eq. 25c	Effective pumping coefficient
R_0	$K_2 \bar{\eta}$	Eq. 12	Characteristic length
τ_0	$\bar{\eta} / P_2^{\text{eff}} $	Eq. 12	Characteristic time
Q_0	$4\pi \bar{\eta}^2 K_2^2 P_2^{\text{eff}} $	Eq. 12	Characteristic volumetric flux
I_0	$4\pi \Lambda_2 \bar{\eta} K_2 P_2^{\text{eff}} / \lambda_2 $	Eq. 12	Characteristic electric current
$\delta_{1,2}$	$P_{1,2}^{\text{eff}} / P_2^{\text{eff}} $	Eq. 18	Dimensionless effective pressure
$\hat{\lambda}$	$\lambda^{\text{eff}} \bar{\eta} K_2 / P_2^{\text{eff}} $	Eq. 18	Dimensionless effective pumping
\hat{v}_2	$(3v_2 - (2(\gamma_2^{\text{app}} + \gamma_0^{\text{app}}) / \bar{\eta})) / (K_2 P_2^{\text{eff}})$	Eq. 18	Dimensionless effective velocity
\hat{Q}	Q^{ext} / Q_0	Below Eq. 13	Dimensionless volumetric flow
\hat{I}	I^{ext} / I_0	Below Eq. 13	Dimensionless electric current
$w_{0,1}$	$j_{0,1} \hat{I} + u_{0,1} \hat{Q}$	Below Eq. 13	Dimensionless external contributions
u_0	$1 - \frac{K_2}{1 - \phi} \left(3\nu_2 - 4\nu_0\nu_4 + \frac{\bar{\kappa}}{\Lambda_2} (3\nu_1 - 4\nu_0\nu_3) \right)$	Eqs. 14 and 15	—
u_1	$\kappa^{\text{eff}} \bar{\eta} K_2^2 / (1 - \phi)$	Eq. 14	—
j_0	$(3\nu_1 - 4\nu_0\nu_3) / (\lambda_2 \bar{\eta} K_2)$	Eqs. 16 and 17	—
j_1	1	Eq. 16	—

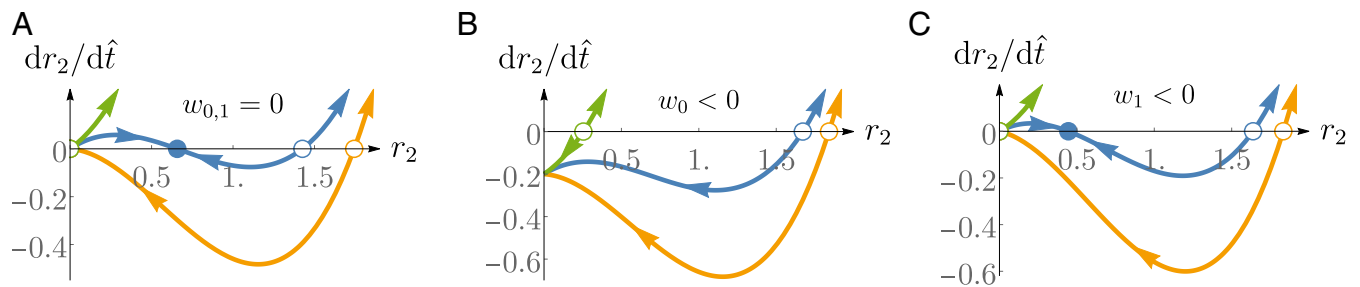


Fig. 4. Phase portraits ($dr_2/d\hat{t}, r_2$) of a spheroid as given by Eq. 13. (A) Example of phase portraits of spheroids that can grow in the absence of a drain. (B) Example of the effect of a negative value of the parameter w_0 on the phase portraits. (C) Example of a negative value of the parameter w_1 on the phase portraits. Parameters (in the order green, blue, orange): $\hat{v}_2 = \{0.5, 0.45, -0.05\}$, $\delta_2 = \{-1, 1, 1\}$, and $\hat{\lambda} = \{2, 1.92, 2.36\}$. Drain parameters: (A) $w_0 = w_1 = 0$, (B) $w_0 = -0.2, w_1 = 0$, and (C) $w_0 = 0, w_1 = -0.1$. Note that we have considered only the numerator of Eq. 13 to draw these phase portraits as the denominator is always positive.

perturbation favors shrinking of the spheroid: The unstable critical radius below which the spheroid shrinks (open circles along the x axis) is shifted to larger values compared to the case without external perturbation shown in Fig. 4A.

We can estimate the critical volumetric flow Q_c needed to change a growing spheroid of size R_2 to a shrinking one. From Eq. 13, we obtain

$$Q_c = -\frac{4\pi K_2 R_2 (\bar{\eta} v_2^{\text{eff}} - P_2^{\text{eff}} R_2 + \lambda^{\text{eff}} R_2^2/4)}{1 + R_2 \kappa^{\text{eff}} K_2/2(1 - \phi) - \beta_n}, \quad [19]$$

where the minus sign indicates that $Q^{\text{ext}} < 0$ is required to induce shrinkage (corresponding to flow out of the drain; Fig. 1B). We obtain a similar expression for the critical electric current that is required to induce shrinkage:

$$I_c = \frac{4\pi R_2 (\bar{\eta} v_2^{\text{eff}} - P_2^{\text{eff}} R_2 + \lambda^{\text{eff}} R_2^2/4)}{-R_2 \lambda_2/(2\Lambda_2) + \bar{\eta} K_2 \beta_j}. \quad [20]$$

Note that $\lambda_2 < 0$ and therefore I_c is positive (corresponding to an electric current into the drain; Fig. 1A). One can therefore always find a value of the externally applied flow or current to turn a growing spheroid into a shrinking one.

State Diagrams of Hydraulic Control of Spheroid Growth. We conclude this section by presenting state diagrams for a spheroid of initial size $R_2(t=0) = R_2^0$ to which a steady external flow Q^{ext} is imposed (Fig. 5). We consider a spheroid with a drain inserted to the center but without lumen. This corresponds to choosing R_1 equal to the outer radius of the drain. For simplicity, we fix $R_1 = 2R_d$, where R_d is the inner drain radius. We again consider spheroids that can grow in the absence of a drain.

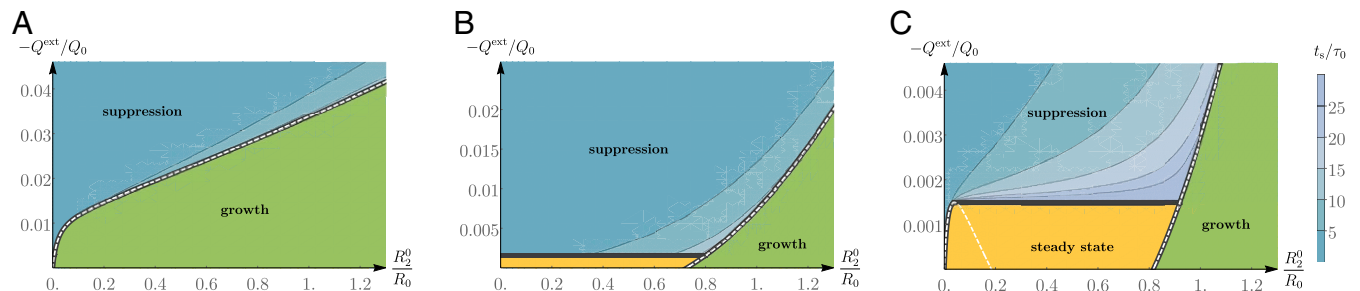


Fig. 5. State diagrams of the long-time growth behavior of the spheroid as a function of its initial radius $R_2(t=0) = R_2^0$ and of the imposed volumetric flux Q^{ext} . The state diagrams correspond to a spheroid which, in the absence of the external flow and for vanishing lumen, is growing for all radii (A), is growing above a critical radius (B), reaches a steady-state radius or grows without bound (C). Shades of blue indicate the time for suppression t_s (main text). Note that all parameters are made dimensionless using the normalization given in Eq. 12. These plots are obtained by solving the dynamics equation Eq. 23b with the parameter values given in Table 3 and with a fixed value of $R_1 = 2R_d$.

Fig. 5A displays the state diagram for the case where the unperturbed spheroid is growing at all radii (corresponding to the green curve in Fig. 4A). Fig. 5B shows the state diagram for a spheroid which is growing above a critical radius (corresponding to the yellow curve in Fig. 4A). Fig. 5C is the state diagram for spheroids that, in the absence of a drain, either reach a steady-state radius or grow without bounds (corresponding to the blue curve in Fig. 4A).

Three different regions exist in the diagrams shown in Fig. 5: 1) a growth region (green), where the imposed flow is not sufficient to arrest growth; note that for increasing values of Q^{ext} , growth slows within this region; 2) a steady-state region (in orange), in which the spheroid reaches a finite size; within this region a larger imposed flow drives the spheroid to smaller steady-state sizes; and 3) a suppression region (in blue) where the spheroid shrinks until it is suppressed. We define that a spheroid has been suppressed when its size becomes smaller than a cutoff size $R_2 = R_1 + h_c$, with $h_c = 10 \mu\text{m}$ a typical cell thickness. In the region of spheroid suppression, the time t_s that it takes for the spheroid to be suppressed is indicated by shades of blue. For fixed initial spheroid size, the time for suppression decreases as the magnitude of the imposed flow increases.

The solid black lines in Fig. 5 indicate boundaries between the different regions. Note that the line separating the steady-state region and the suppression region depends on the cutoff h_c . The dashed white line indicates the nullcline $dr_2/d\hat{t} = 0$ and gives the critical value of the external flow Q_c required to switch from a growing to a shrinking spheroid (given by Eq. 19 for $R_1 = 0$). Note that below the nullcline the spheroids grow and above the nullcline they shrink. Note that a similar analysis can be performed in the case of an external current I^{ext} . In this case the same behaviors are found: growth, suppression, and

steady-state regions which depend on the initial spheroid size and on the magnitude of the imposed electric current.

Conclusion

Using a coarse-grained description of a tissue as an active material capable of exerting mechanical stresses, transporting ions, and pumping fluid, we have proposed a technique to perturb tissues electrically and hydraulically. This technique relies on an imposed fluid flow or electric current source using a micro-metric drain or electrode which can turn a growing spheroid to shrinkage.

Our theory allows us to estimate orders of magnitude of the external flow and current that are required to suppress an initially growing spheroid. For small spheroids with radius of about 100 μm , we show for instance that an electric current of a few nanoamperes or a volumetric flow of about $10^3 \mu\text{m}^3/\text{s}$ could have a significant impact on the spheroid state (Fig. 6 in Appendix E). It means that even a passive drain connecting the inner part of the spheroid to the surrounding medium could already alter growth of small spheroids. For larger aggregates, connecting the drain to a pump leads to spheroid suppression for any size, provided that the imposed external flow is sufficiently strong and maintained long enough (for instance, for spheroids of a few millimeters in radius an electric current of tens to hundreds of nanoamperes or a volumetric flow of about 10^4 to $10^5 \mu\text{m}^3/\text{s}$ could be sufficient; Fig. 6). Spheroid suppression can be obtained also by imposing an electric current, and both fluid flow and electric current application could be combined to accelerate the process.

Our approach has also allowed us to characterize the different long-time states of a growing spheroid subject to an external perturbation. Depending on the magnitude of the hydraulic or electrical field, we have shown that the growth can be either slowed down or arrested (Fig. 5). In the latter case, the arrest of growth can lead to a steady state or even to the shrinking and eventually the suppression of the spheroid. If the magnitude of the external field is above a threshold value, the suppression of the spheroid is always achieved, and stronger magnitudes then accelerate the process. Our coarse-grained description is generic, as it depends on effective tissue parameters but is insensitive to many details. Our approach does not distinguish between a suppressed state where all cells have disappeared and a state where a few cells still remain. This distinction depends on cellular details that we are not considering here.

We have also shown how different protocols can be used to obtain this suppression: For instance, a longer perturbation at a weaker magnitude is slower but as efficient as a shorter but stronger one; decreasing the magnitude of the flow or electric current as the size of the spheroid decreases can also be used to suppress a spheroid (see Fig. 2 for examples). Interestingly, protocols that could lead to spheroid suppression need to be carried out over a sufficiently long time period. We estimate that the slow, steady flow or electric current that mediates the progressive suppression of a spheroid has to be maintained over days or

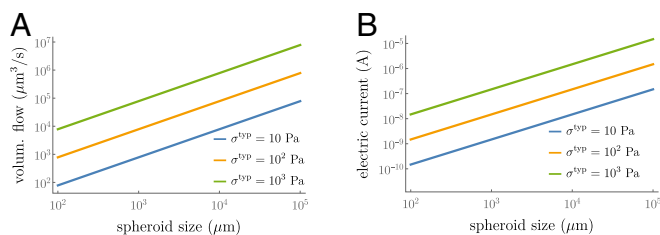


Fig. 6. Orders of magnitude of the external fields required to observe a significant change in the spheroid dynamics as a function of their size as given by Eqs. 10 and 11. (A) Volumetric flow Q^{ext} . (B) Electric current I^{ext} .

weeks. This is in contrast with typical cancer ablation techniques (35), for which treatments are brief and intense. Radiofrequency ablation for instance requires the application of an alternating current with high frequency (up to 500 kHz) and high voltage (up to several kilovolts) to heat the tissue (46). Similarly, irreversible electroperoration ablations use microsecond pulses of high electric potential (up to 3 kV) (33).

At the time scales considered in this article, the case of a tumor is more complex than that of a spheroid. First, one has to compare the effect of the flux on the tumor with that on the surrounding healthy tissue. This question will be addressed in future work. Second, we have not explicitly considered the role of nutrient and oxygen transport. The larger division rate at the surface that we consider (Eq. 9) could be related for example to the oxygen gradient that has been observed experimentally (47). Furthermore, nutrient transport has been shown to influence growth in other contexts (48). Moreover, the geometry of spheroids could be more complex than a simple sphere, and a natural expansion of this current work will be the study of the stability of the spherical shape during the growth or shrinkage processes (49). Last, during the suppression process, the escape of metastatic cells should be prevented. The signs of stresses that we find here correspond to flows toward the sphere center. This suggests that escape of material might be hampered by imposed flows or currents, thereby providing a barrier against the escape of cells.

Our work is based on generic mechanical, hydraulic, and electrical properties of tissues. It thus paves the way for experimental methods to control spheroid size. If applied to cancerous tissues, it could provide a means to influence cancerous tumor growth, arrest their proliferation, and even suppress these tumors. Indeed, the hydraulic and electrical perturbations we have proposed here do not rely on identifying and hindering specific chemical pathways that are characteristic of cancerous tissue (50), but rather on general, physical responses to an external field.

Appendix A. Constitutive Equations for Permeation and for the Electric Current Density

Based on the symmetry of the system and assuming linear response, the constitutive equation for the internal force density reads

$$f_{\alpha} = -\tilde{\kappa}(v_{\alpha}^c - v_{\alpha}^f) - \tilde{\kappa}' q_{\alpha\beta}(v_{\beta}^c - v_{\beta}^f) + \tilde{\lambda}_1 p_{\alpha} + \tilde{\lambda}'_1 q_{\alpha\beta} p_{\beta} + \tilde{\lambda}_2 E_{\alpha} + \tilde{\lambda}'_2 q_{\alpha\beta} E_{\beta} + \tilde{\lambda}_3 p_{\alpha} \partial_{\beta} p_{\beta} + \tilde{\lambda}'_3 p_{\beta} \partial_{\beta} p_{\alpha}. \quad [21]$$

Note that the term $p_{\alpha} \partial_{\beta} p_{\alpha} = \partial_{\beta}(p_{\alpha} p_{\alpha})/2$ vanishes as the polarity is a unit vector. Moreover, in the main text we consider a system with spherical symmetry and the polarity is assumed to be radial $\mathbf{p} = \mathbf{e}_r$, such that the nematic order parameter $q_{\alpha\beta} = p_{\alpha} p_{\beta} - (1/3)\delta_{\alpha\beta}$ is diagonal in spherical coordinates with $q_{rr} = 2/3$, $q_{\theta\theta} = -1/3$, $q_{\varphi\varphi} = -1/3$. As a consequence, and since $p_{\beta} \partial_{\beta} p_{\alpha} = 0$ for $\mathbf{p} = \mathbf{e}_r$, we obtain Eq. 3 in the main text with $\tilde{\kappa} = \tilde{\kappa} + 2\tilde{\kappa}'/3$, $\lambda_1 = \tilde{\lambda}_1 + 2\tilde{\lambda}'_1/3$, $\lambda_2 = \tilde{\lambda}_2 + 2\tilde{\lambda}'_2/3$, and $\lambda_3 = \tilde{\lambda}_3$.

Similarly, the general form for the electric current density constitutive equation reads

$$j_{\alpha} = -k(v_{\alpha}^c - v_{\alpha}^f) - k' q_{\alpha\beta}(v_{\beta}^c - v_{\beta}^f) + \tilde{\Lambda}_1 p_{\alpha} + \tilde{\Lambda}'_1 q_{\alpha\beta} p_{\beta} + \tilde{\Lambda}_2 E_{\alpha} + \tilde{\Lambda}'_2 q_{\alpha\beta} E_{\beta} + \tilde{\Lambda}_3 p_{\alpha} \partial_{\beta} p_{\beta} + \tilde{\Lambda}'_3 p_{\beta} \partial_{\beta} p_{\alpha} \quad [22]$$

and it yields Eq. 4 in the main text with $\bar{\kappa} = k + 2k'/3$, $\Lambda_1 = \tilde{\Lambda}_1 + 2\tilde{\Lambda}'_1/3$, $\Lambda_2 = \tilde{\Lambda}_2 + 2\tilde{\Lambda}'_2/3$, and $\Lambda_3 = \tilde{\Lambda}_3$ for a system with spherical symmetry and with $\mathbf{p} = \mathbf{e}_r$. For a system without spherical symmetry or with $\mathbf{p} \neq \mathbf{e}_r$, Eqs. 21 and 22 must be used.

Appendix B. Dynamics of a Spheroid in the Presence of a Drain

The dynamical equations for the spheroid and lumen radii are obtained by first considering force balance Eq. 1 in spherical coordinates with boundary conditions Eq. 9. This yields the velocity profiles of cells and interstitial fluid. A differential equation for the radii dynamics $R_{1,2}(t)$ is then obtained from the normal stress balance Eq. 7 together with the permeation conditions at the boundary given by Eq. 8. Details on this computation in the case without a drain can be found in ref. 27. Including the external fields due to the presence of the drain, we obtain the following coupled differential equations for the dynamics of the radii r_1 and r_2 of the lumen and of the spheroid:

$$\begin{aligned} \chi \frac{dr_1}{dt} + \frac{3(\hat{V}_1 + dr_1/d\hat{t})r_1^2}{r_2^3 - r_1^3} + \frac{3(\hat{V}_2 - dr_2/d\hat{t})r_2^2}{r_2^3 - r_1^3} &= \delta_1 \\ - \frac{2}{r_1} \left(\hat{\gamma}_1 - \hat{\gamma}_0 \frac{r_1(r_1 + r_2)}{r_1^2 + r_2^2 + r_1 r_2} \right) \\ + (u_1 \hat{Q} + \hat{I}) \frac{1}{r_1} \frac{(r_2 - r_1)(r_1 + 2r_2)}{2(r_1^2 + r_2^2 + r_1 r_2)} \\ + \frac{1}{r_1^2} \left(\chi \hat{Q} + u_2 \hat{Q} + j_2 \hat{I} - (u_3 \hat{Q} + j_3 \hat{I}) \frac{(r_2 - r_1)(2r_1 + r_2)}{r_1^2 + r_2^2 + r_1 r_2} \right) \\ + \hat{\lambda}(r_2 - r_1) \frac{r_1^2 + 2r_1 r_2 + 3r_2^2}{4(r_1^2 + r_2^2 + r_1 r_2)}, \end{aligned} \quad [23a]$$

$$\begin{aligned} - \frac{dr_2}{dt} + \frac{3(\hat{V}_1 + dr_1/d\hat{t})r_1^2}{r_2^3 - r_1^3} + \frac{3(\hat{V}_2 - dr_2/d\hat{t})r_2^2}{r_2^3 - r_1^3} &= \delta_2 \\ + \frac{2}{r_2} \left(\hat{\gamma}_2 + \hat{\gamma}_0 \frac{r_2(r_1 + r_2)}{r_1^2 + r_2^2 + r_1 r_2} \right) \\ - (u_1 \hat{Q} + \hat{I}) \frac{1}{r_2} \frac{(r_2 - r_1)(2r_1 + r_2)}{2(r_1^2 + r_2^2 + r_1 r_2)} \\ + \frac{1}{r_2^2} \left(-\hat{Q} + u_2 \hat{Q} + j_2 \hat{I} + (u_3 \hat{Q} + j_3 \hat{I}) \frac{(r_2 - r_1)(r_1 + 2r_2)}{r_1^2 + r_2^2 + r_1 r_2} \right) \\ - \hat{\lambda}(r_2 - r_1) \frac{3r_1^2 + 2r_1 r_2 + r_2^2}{4(r_1^2 + r_2^2 + r_1 r_2)}, \end{aligned} \quad [23b]$$

where we have introduced dimensionless radii $r_i(\hat{t}) = R_i(t)/R_0$ with $R_0 = K_2 \bar{\eta}$ and a dimensionless time $\hat{t} = t/\tau_0$ with $\tau_0 = \bar{\eta}/|P_2^{\text{eff}}|$. We have also introduced the dimensionless parameters

$$\begin{aligned} \hat{\gamma}_{0,1,2} &= \frac{\gamma_{0,1,2}^{\text{app}}}{\bar{\eta} K_2 |P_2^{\text{eff}}|}, \quad \hat{V}_{1,2} = \frac{v_{1,2}}{K_2 |P_2^{\text{eff}}|}, \quad \delta_{1,2} = \frac{P_{1,2}^{\text{eff}}}{|P_2^{\text{eff}}|} \\ \hat{\lambda} &= \frac{\lambda^{\text{eff}} \bar{\eta} K_2}{|P_2^{\text{eff}}|}, \quad \chi = \frac{K_2}{K_1}, \end{aligned} \quad [24]$$

where the effective parameters are defined as

$$\begin{aligned} P_{1,2}^{\text{eff}} &= \Pi_{1,2}^{\text{ext}} - P_h^c - \frac{J_{p,1,2}}{K_{1,2}} \\ &- \frac{2}{3} \left(\zeta_0 \nu_0 + \lambda_3 + \frac{\Lambda_3 \lambda_2 + (3\nu_1/2 - 2\nu_0 \nu_3) \Lambda_1}{\Lambda_2} \right), \end{aligned} \quad [25a]$$

$$\begin{aligned} \gamma_{1,2}^{\text{app}} &= \gamma_{1,2} \mp 4\nu_3 \Lambda_3 / \Lambda_2, \\ \gamma_0^{\text{app}} &= (3\nu_1/2 - 2(2 + \nu_0) \nu_3) (\Lambda_3 / \Lambda_2), \end{aligned} \quad [25b]$$

$$\lambda^{\text{eff}} = \lambda_1 - \Lambda_1 \lambda_2 / \Lambda_2. \quad [25c]$$

In addition to these terms, the presence of external flows and currents gives new dimensionless contributions which are defined as

$$\hat{Q} = \frac{Q^{\text{ext}}}{4\pi \bar{\eta}^2 K_2^3 |P_2^{\text{eff}}|}, \quad \hat{I} = \frac{I^{\text{ext}} |\lambda_2|}{4\pi \Lambda_2 \bar{\eta} K_2 |P_2^{\text{eff}}|}, \quad u_1 = \frac{\kappa^{\text{eff}} \bar{\eta} K_2^2}{1 - \phi}, \quad [26a]$$

$$u_2 = \frac{K_2}{1 - \phi} \left(\nu_2 - 4(\nu_0 - 1) \nu_4 / 3 - \frac{\bar{\kappa}}{\Lambda_2} (4(\nu_0 - 1) \nu_3 / 3 - \nu_1) \right), \quad [26b]$$

$$u_3 = \frac{K_2}{1 - \phi} \left(\nu_2 - 2(1 + 2\nu_0) \nu_4 / 3 - \frac{\bar{\kappa}}{\Lambda_2} (2(1 + 2\nu_0) \nu_3 / 3 - \nu_1) \right), \quad [26c]$$

$$j_2 = \frac{4(\nu_0 - 1) \nu_3 / 3 - \nu_1}{\lambda_2 \bar{\eta} K_2}, \quad j_3 = \frac{2(1 + 2\nu_0) \nu_3 / 3 - \nu_1}{\lambda_2 \bar{\eta} K_2}, \quad [26d]$$

where $\kappa^{\text{eff}} = \kappa - \bar{\kappa} \lambda_2 / \Lambda_2$ is an effective permeation coefficient.

To understand the effect of an external flux and an external electric current on the spheroid size, we can rewrite Eq. 23b in the form of an equation for the dynamics of the spheroid volume $V = 4\pi R_2^3/3$. To simplify the analysis, we take a vanishing lumen size ($r_1 = 0$) and we consider only the term that comes from the externally imposed current and flux. It yields

$$\begin{aligned} \frac{dV}{dt} &= Q^{\text{ext}} + Q^{\text{ext}} \frac{K_2 \kappa^{\text{eff}} R_2}{1 - \phi} + I^{\text{ext}} \frac{K_2 \lambda_2 R_2}{\Lambda_2} - Q^{\text{ext}} (u_2 + 2u_3) \\ &- I^{\text{ext}} \frac{\lambda_2 K_2}{\Lambda_2} (j_2 + 2j_3), \end{aligned} \quad [27]$$

where we have reintroduced dimensional quantities to simplify the following discussion. The first term on the right-hand side of Eq. 27 is a direct consequence of imposing an external flow: Since the tissue is incompressible, an imposed flow from the outside to the inside of the spheroid ($Q^{\text{ext}} > 0$) provokes an increase in volume, while an imposed flow from the inside of the spheroid to the outside ($Q^{\text{ext}} < 0$) shrinks its size. The second term is a signature of the finite permeability of the tissue, which acts as a porous medium, and the effect is therefore proportional to the ratio of the surface permeability K_2 and the bulk permeability $(\kappa^{\text{eff}} R_2 / (1 - \phi))^{-1}$. The third term accounts for a similar phenomenon to the second one but due to the electroosmotically generated flow due to the imposed current. The two last terms correspond to bioelectric and biohydraulic contributions: Indeed, the terms $u_{2,3}$ and $j_{2,3}$ involve the parameters ν_i and thus stem from the coupling between the electric field (or the interstitial fluid flow) and the cell polarity that appears in the cell stress.

In the case of a small lumen compared to the spheroid size, one can consider the limit $R_1 \ll R_2$ to describe the spheroid dynamics. In this limit, we can in particular obtain from Eq. 23b an equation for the dynamics of r_2 only, which is given by Eq. 13 in the main text. The parameters $\beta_{i,j}$ introduced in the main text read $\beta_u = u_2 + 2u_3$ and $\beta_j = j_2 + 2j_3$.

Appendix C. Imposed Pressure and Electric Potential Difference

The main text describes the spheroid size control when an external volumetric flow Q^{ext} or an external current I^{ext} is imposed. Alternatively, a pressure difference or an electric potential difference can be imposed between the outer part of the drain and the outer layer of the spheroid, which can be considered as the

conjugate ensemble. Here we provide the additional equations required to discuss this situation.

Imposed Pressure Difference. To compute the pressure difference $\Delta P = P^d - P_2^{\text{ext}}$, we decompose it as $\Delta P = \Delta P_{d1} + \Delta P_{12}$, where $\Delta P_{12} = P_1^{\text{ext}} - P_2^{\text{ext}}$ is the pressure difference across the spheroid and $\Delta P_{d1} = P^d - P_1^{\text{ext}}$ is the pressure difference between the outer part of the drain and the lumen and reads

$$\Delta P_{d1} = Q^{\text{ext}}(K_d + K_{\text{lum}}), \quad [28]$$

where $K_{d,\text{lum}}$ are the hydraulic resistances of the drain and of the lumen, respectively. The drain hydraulic resistance is given by $K_d = \frac{8\eta^f L_d}{\pi R_d^4}$, where $\eta^f \simeq 10 \text{ mPa}\cdot\text{s}$ is the viscosity of the interstitial fluid, and L_d, R_d are the width and the radius of the drain.

The pressure difference $\Delta P_{12} = P_1^{\text{ext}} - P_2^{\text{ext}}$ can then be computed using Eq. 8. In dimensionless form, it reads

$$\Delta \hat{P}_{12} = \Delta \hat{\Pi} + \hat{Q} \left(\frac{\chi}{r_1^2} + \frac{1}{r_2^2} \right) - (\chi \hat{r}_1 + \hat{r}_2) - \Delta \hat{P}^f, \quad [29]$$

where we use the short-hand notation $\hat{r}_{1,2} = dr_{1,2}/d\hat{t}$ and we have defined

$$\begin{aligned} \Delta \hat{P}_{12} &= \frac{P_1^{\text{ext}} - P_2^{\text{ext}}}{|P_2^{\text{eff}}|}, \quad \Delta \hat{P}^f = \frac{P^f(r_1) - P^f(r_2)}{|P_2^{\text{eff}}|} \\ \Delta \hat{\Pi} &= \frac{\Pi_1 - \Pi_2 - (J_1/K_1 - J_2/K_2)}{|P_2^{\text{eff}}|}. \end{aligned} \quad [30]$$

The dimensionless quantities P_2^{eff} , $r_{1,2}$, χ , and \hat{Q} have been introduced in *Appendix B*. To compute the interstitial pressure difference $\Delta P^f = P^f(R_1) - P^f(R_2)$, one can use the identity $\Delta P^f = -\int_{R_1}^{R_2} dr \partial_r P^f(r)$. Indeed, the gradient of the interstitial fluid pressure can be obtained using force balance Eq. 1b and the constitutive equations Eqs. 3 and 4. It reads

$$\partial_{\hat{r}} \hat{P}^f(\hat{r}) = u_1 \hat{v}(\hat{r}) - \hat{\lambda} - \frac{2\hat{\lambda}_3}{\hat{r}} - \frac{u_1 \hat{Q} + \hat{I}}{\hat{r}^2}, \quad [31]$$

where we have introduced, in addition to the dimensionless quantities already introduced in *Appendix B*, the dimensionless radius $\hat{r} = r/R_0$, cell velocity $\hat{v} = v_r^c/(K_2|P_2^{\text{eff}}|)$, and interstitial fluid pressure $\hat{P}^f = P^f/|P_2^{\text{eff}}|$ and the dimensionless parameter $\hat{\lambda}_3 = (\lambda_3 - \Lambda_3 \lambda_2/\Lambda_2)/|P_2^{\text{eff}}|$. The dimensionless cell velocity appearing in Eq. 31 can be cast into the form

$$\hat{v}(\hat{r}) = \frac{a}{\hat{r}^2} + \frac{b}{\hat{r}} + c + d\hat{r} - \frac{\hat{\lambda}}{4}\hat{r}^2, \quad [32]$$

where we have introduced the \hat{r} -independent factors

$$\begin{aligned} a &= \frac{r_1^2 r_2^2}{r_{12}^2} (r_1(\tilde{v}_2 - \hat{r}_2) + r_2(\tilde{v}_1 + \hat{r}_1)) - \frac{\hat{\lambda} r_1^3 r_2^3}{4r_{12}^2} \\ &+ \frac{\hat{Q} r_1 r_2}{r_{12}^2} \left(u_3(r_1 + r_2) - \frac{u_1 r_1 r_2}{2} \right) \\ &+ \frac{\hat{I} r_1 r_2}{r_{12}^2} \left(j_3(r_1 + r_2) - \frac{r_1 r_2}{2} \right), \end{aligned} \quad [33a]$$

$$b = -u_3 \hat{Q} - j_3 \hat{I}, \quad c = \frac{u_1 \hat{Q}}{2} + \frac{\hat{I}}{2} - \frac{2\hat{\gamma}_0}{3}, \quad [33b]$$

$$\begin{aligned} d &= \frac{\hat{\lambda}}{4} \left(r_2 + \frac{r_1^3}{r_{12}^2} \right) - \frac{(\tilde{v}_1 + \hat{r}_1)r_1^2 + (\tilde{v}_2 - \hat{r}_2)r_2^2}{r_2^3 - r_1^3} \\ &+ \frac{\hat{Q}}{r_{12}^2} \left(u_3 - \frac{u_1(r_1 + r_2)}{2} \right) + \frac{\hat{I}}{r_{12}^2} \left(j_3 - \frac{r_1 + r_2}{2} \right), \end{aligned} \quad [33c]$$

and the dimensionless quantities (in addition to those already introduced in *Appendix B*)

$$r_{12}^2 = r_1^2 + r_2^2 + r_1 r_2, \quad \tilde{v}_{1,2} = \hat{V}_{1,2} \mp 2\hat{\gamma}_0/3. \quad [34]$$

The dimensionless pressure difference Eq. 29 can therefore be computed in terms of the dimensionless parameters of the model, although its expression is lengthy. One can, however, expand this expression in the limit of a thin spheroid (i.e., $R_1 \sim R_2$), yielding the expression

$$\Delta \hat{P}_{12} = \frac{1}{|P_2^{\text{eff}}|} \frac{2(\gamma_1 + \gamma_2)}{R_2} + (r_2 - r_1)A + \mathcal{O}((r_2 - r_1)^2), \quad [35]$$

where the first term on the right-hand side is the Laplace contribution in the limit $R_1 \rightarrow R_2$ and the dimensionless coefficient A represents the first correction in terms of the thickness $R_2 - R_1$ and reads

$$\begin{aligned} A &= \frac{u_1 (\hat{V}_2 - \chi \hat{V}_1 - \Delta \hat{\Pi})}{1 + \chi} + \frac{2}{r_2} \left(\frac{u_1(\hat{\gamma}_1 + \hat{\gamma}_2)}{1 + \chi} + \hat{\lambda}_3 \right) + \frac{2\hat{\gamma}_1}{r_2^2} \\ &+ \frac{\hat{Q}}{r_2^2} \left(\frac{2(u_2 - u_3)}{r_2} - u_1 \right) + \frac{2\hat{I}(j_3 - j_2)}{r_2^3}. \end{aligned} \quad [36]$$

Imposed Electric Potential Difference. Similar to the pressure, one can compute the electric potential difference $\Delta U = U^d - U_2^{\text{ext}}$ by decomposing it as $\Delta U = \Delta U_{d1} + \Delta U_{12}$ with $\Delta U_{12} = U_1^{\text{ext}} - U_2^{\text{ext}}$ the electric potential difference across the spheroid and with $\Delta U_{d1} = U^d - U_1^{\text{ext}}$ the electric potential difference across the drain. This difference reads

$$\Delta U_{d1} = I^{\text{ext}}(1/G_d + 1/G_{\text{lum}}), \quad [37]$$

where $G_{d,\text{lum}}$ are the electrical conductances of the drain and of the lumen. The electrical conductance of the drain is computed as $1/G_d = \frac{\rho^f L_d}{\pi R_d^3}$, where $\rho^f \simeq 1 \Omega\cdot\text{m}$ is the resistivity of the interstitial fluid (taken to be that of salted water, whose conductivity is about $G \simeq 1 \text{ S/m}$).

The electric potential difference across the spheroid ΔU_{12} can then be computed in the following way. From Eq. 4, an expression for the electric field can be obtained, which can then be used to compute the electric potential difference using the identity $\Delta U_{12} = \int_{R_1}^{R_2} dr E(r)$. Defining the dimensionless electric field $\hat{E} = E_r \bar{\eta} K_2 \lambda_2 / |P_2^{\text{eff}}|$, we have

$$\hat{E}(\hat{r}) = -\hat{\Lambda}_1 - \bar{u}_1 \hat{v}(\hat{r}) - \frac{2\hat{\Lambda}_3}{\hat{r}} - \frac{\bar{u}_1 \hat{Q} - \hat{I}}{\hat{r}^2}, \quad [38]$$

where we have defined the dimensionless parameters

$$\bar{u}_1 = \frac{\bar{\eta} K_2^2 \bar{\kappa} \lambda_2}{(1 - \phi) \Lambda_2}, \quad \hat{\Lambda}_1 = \frac{\bar{\eta} K_2 \lambda_2 \Lambda_1}{\Lambda_2 |P_2^{\text{eff}}|}, \quad \hat{\Lambda}_3 = \frac{\lambda_2 \Lambda_3}{\Lambda_2 |P_2^{\text{eff}}|}, \quad [39]$$

and where the expression for the dimensionless cell velocity \hat{v} is displayed in Eq. 32.

Appendix D. Numerical Solution to the Dynamics Equations

Figs. 2 and 3 of the main text have been obtained by solving the coupled equations Eqs. 23a and 23b using the dimensionless parameters given in Table 3. The presence of the drain provides

Table 3. Dimensionless parameter values used for plotting the figures

Figure	Parameter values													
	δ_1	δ_2	χ	$\hat{\gamma}_0$	$\hat{\gamma}_1$	$\hat{\gamma}_2$	$\hat{\lambda}$	\hat{V}_1	\hat{V}_2	u_1	u_2	u_3	j_2	j_3
Fig. 2	0.1	-1	1	0	-0.05	0.15	0	1	0.1	10^2	0	0	0	0
Fig. 3	0.3	-1	1	0	0.15	0.15	0.2	0.1	0.5	10^2	0	0	0	0
Fig. 5A	—	-1	1	0	—	0.15	0.5	—	0.3	10^2	0	0	0	0
Fig. 5B	—	1	1	-0.15	—	0.15	5.5	—	0	10^2	0	0	0	0
Fig. 5C	—	1	1	0	—	0.15	4	—	0.15	10^2	0	0	0	0

These parameters are defined in Eqs. 24–26 in Appendix B.

a natural cutoff for the (dimensionless) lumen radius r_1 : If at a time t_0 the inner radius $R_1(t)$ reaches the size of the outer drain radius, which we assume to be equal to $2R_d = 10 \mu\text{m}$, we fix its (dimensionless) value to be $r_1(t > t_0) = 2R_d/R_0$ and solve only Eq. 23b with this fixed value for r_1 . If the external flow is stopped at a subsequent time $t_1 > t_0$ (see Fig. 2F of the main text for instance), we restart solving simultaneously Eqs. 23a and 23b, taking $r_1(t_1) = 2R_d/R_0$ as the initial condition.

Fig. 5 of the main text has been obtained by taking $r_1 = 2R_d/R_0$ and solving numerically Eq. 23b using the dimensionless parameters given in Table 3, with the initial condition $R_2(t=0) = R_2^0$ and a given imposed external flux Q^{ext} . If a spheroid reaches a cutoff size $R_2 = 2R_d + h_c$ after a time $t = t_s$, it is classified as “suppressed” and its suppression time t_s is used to produce the color coding.

Appendix E. Estimates for Parameter Values

Estimation of the phenomenological parameters appearing in the continuum model is crucial for our analysis. Some of these parameters, such as the cell shear and bulk viscosities, the tissue surface tension, and the bulk permeability for instance, have already been estimated in experiments. For most of the remaining parameters, experimental values are not yet available and we have used order-of-magnitude estimations (see refs. 26 and 27 for details). We provide parameter values and references for these values in Table 1.

Data Availability. There are no data underlying this work.

ACKNOWLEDGMENTS. We thank C. T. Lim and T. B. Saw for useful discussions.

- J. B. A. Green, J. Sharpe, Positional information and reaction-diffusion: Two big ideas in developmental biology combine. *Development* **142**, 1203–1211 (2015).
- C. J. Miller, L. A. Davidson, The interplay between cell signalling and mechanics in developmental processes. *Nat. Rev. Genet.* **14**, 733–744 (2013).
- T. Mammoto, A. Mammoto, D. E. Ingber, Mechanobiology and developmental control. *Annu. Rev. Cell Dev. Biol.* **29**, 27–61 (2013).
- B. Ladoux, R. M. Mège, Mechanobiology of collective cell behaviours. *Nat. Rev. Mol. Cell Biol.* **18**, 743–757 (2017).
- T. Ruiz-Herrero, K. Alessandri, B. V. Gurchenkov, P. Nassoy, L. Mahadevan, Organ size control via hydraulically gated oscillations. *Development* **144**, 4422–4427 (2017).
- J. G. Dumortier et al., Hydraulic fracturing and active coarsening position the lumen of the mouse blastocyst. *Science* **365**, 465–468 (2019).
- M. Levin, C. J. Martyniuk, The bioelectric code: An ancient computational medium for dynamic control of growth and form. *Biosystems* **164**, 76–93 (2018).
- B. B. Silver, C. M. Nelson, The bioelectric code: Reprogramming cancer and aging from the interface of mechanical and chemical microenvironments. *Front. Cell Dev. Biol.* **6**, 21 (2018).
- C. J. Chan et al., Hydraulic control of mammalian embryo size and cell fate. *Nature* **571**, 112–116 (2019).
- C. Fütterer, C. Colombo, F. Jülicher, A. Ott, Morphogenetic oscillations during symmetry breaking of regenerating *Hydra vulgaris* cells. *Europhys. Lett.* **64**, 137–143 (2003).
- W. Roux, Über die morphologische polarisation von eiern und embryonen durch den elektrischen strom. *Math. Naturwiss. Kl.* **101**, 27–228 (1892).
- C. D. McCaig, A. M. Rajnicek, B. Song, M. Zhao, Controlling cell behavior electrically: Current views and future potential. *Physiol. Rev.* **85**, 943–978 (2005).
- K. A. McLaughlin, M. Levin, Bioelectric signaling in regeneration: Mechanisms of ionic controls of growth and form. *Dev. Biol.* **433**, 177–189 (2018).
- H. Watanabe, T. Fujisawa, T. W. Holstein, Cnidarians and the evolutionary origin of the nervous system: Cnidarian nervous system. *Dev. Growth Differ.* **51**, 167–183 (2009).
- E. Braun, H. Ori, Electric-induced reversal of morphogenesis in *Hydra*. *Biophys. J.* **117**, 1514–1523 (2019).
- F. Martín-Belmonte et al., Cell-polarity dynamics controls the mechanism of lumen formation in epithelial morphogenesis. *Curr. Biol.* **18**, 507–513 (2008).
- J. Debnath et al., The role of apoptosis in creating and maintaining luminal space within normal and oncogene-expressing mammary acini. *Cell* **111**, 29–40 (2002).
- M. Huch, B. K. Koo, Modeling mouse and human development using organoid cultures. *Development* **142**, 3113–3125 (2015).
- S. Dahl-Jensen, A. Grapin-Botton, The physics of organoids: A biophysical approach to understanding organogenesis. *Development* **144**, 946–951 (2017).
- J. Alcaraz et al., Laminin and biomimetic extracellular elasticity enhance functional differentiation in mammary epithelia. *EMBO J.* **27**, 2829–2838 (2008).
- F. Montel et al., Stress clamp experiments on multicellular tumor spheroids. *Phys. Rev. Lett.* **107**, 188102 (2011).
- M. Delarue et al., Mechanical control of cell flow in multicellular spheroids. *Phys. Rev. Lett.* **110**, 138103 (2013).
- K. Kruse, J. F. Joanny, F. Jülicher, J. Prost, K. Sekimoto, Generic theory of active polar gels: A paradigm for cytoskeletal dynamics. *Eur. Phys. J. E* **16**, 5–16 (2005).
- M. C. Marchetti et al., Hydrodynamics of soft active matter. *Rev. Mod. Phys.* **85**, 1143–1189 (2013).
- J. Ranft et al., Fluidization of tissues by cell division and apoptosis. *Proc. Natl. Acad. Sci. U.S.A.* **107**, 20863–20868 (2010).
- N. Sarkar, J. Prost, F. Jülicher, Field induced cell proliferation and death in a model epithelium. *New J. Phys.* **21**, 043035 (2019).
- C. Duclut, N. Sarkar, J. Prost, F. Jülicher, Fluid pumping and active flexoelectricity can promote lumen nucleation in cell assemblies. *Proc. Natl. Acad. Sci. U.S.A.* **116**, 19264–19273 (2019).
- A. M. Scott, J. D. Wolchok, L. J. Old, Antibody therapy of cancer. *Nat. Rev. Canc.* **12**, 278–287 (2012).
- R. Baskar, K. A. Lee, R. Yeo, K. W. Yeoh, Cancer and radiation therapy: Current advances and future directions. *Int. J. Med. Sci.* **9**, 193–199 (2012).
- O. Al-Bataineh, J. Jenne, P. Huber, Clinical and future applications of high intensity focused ultrasound in cancer. *Canc. Treat. Rev.* **38**, 346–353 (2012).
- D. R. Mittelstein et al., Selective ablation of cancer cells with low intensity pulsed ultrasound. *Appl. Phys. Lett.* **116**, 013701 (2020).
- A. Tijore et al., Ultrasound-mediated mechanical forces selectively kill tumor cells. bioRxiv: [Preprint] (2020). <https://doi.org/10.1101/2020.10.09.332726> (Accessed 9 October 2020).
- L. Miller, J. Leor, B. Rubinsky, Cancer cells ablation with irreversible electroporation. *Technol. Canc. Res. Treat.* **4**, 699–705 (2005).
- N. R. Perkons et al., Electrolytic ablation enables cancer cell targeting through pH modulation. *Commun. Biol.* **1**, 48 (2018).
- E. M. Knavel, C. L. Brace, Tumor ablation: Common modalities and general practices. *Tech. Vasc. Interv. Radiol.* **16**, 192–200 (2013).
- J. Ranft, J. Prost, F. Jülicher, J. F. Joanny, Tissue dynamics with permeation. *Eur. Phys. J. E* **35**, 46 (2012).
- M. E. Dolega et al., Cell-like pressure sensors reveal increase of mechanical stress towards the core of multicellular spheroids under compression. *Nat. Commun.* **8**, 14056 (2017).
- M. Basan, T. Risler, J. F. Joanny, X. Sastre-Garau, J. Prost, Homeostatic competition drives tumor growth and metastasis nucleation. *HFSP J.* **3**, 265–272 (2009).
- R. Simha, S. Ramaswamy, Hydrodynamic fluctuations and instabilities in ordered suspensions of self-propelled particles. *Phys. Rev. Lett.* **89**, 058101 (2002).
- T. Bittig, O. Wartlick, A. Kicheva, M. González-Gaitán, F. Jülicher, Dynamics of anisotropic tissue growth. *New J. Phys.* **10**, 063001 (2008).
- D. J. Blackiston, K. A. McLaughlin, M. Levin, Bioelectric controls of cell proliferation: Ion channels, membrane voltage and the cell cycle. *Cell Cycle* **8**, 3527–3536 (2009).

42. H. P. G. Darcy, *Les Fontaines Publiques de La Ville de Dijon* (Dalmont, Paris, France, 1856).
43. S. Ramaswamy, J. Toner, J. Prost, Nonequilibrium fluctuations, traveling waves, and instabilities in active membranes. *Phys. Rev. Lett.* **84**, 3494–3497 (2000).
44. B. J. Kirby, *Micro- and Nanoscale Fluid Mechanics* (Cambridge University Press, Cambridge, England, 2013).
45. M. Delarue, J. F. Joanny, F. Jülicher, J. Prost, Stress distributions and cell flows in a growing cell aggregate. *Interf. Focus* **4**, 20140033 (2014).
46. A. Erez, A. Shitzer, Controlled destruction and temperature distributions in biological tissues subjected to monoactive electrocoagulation. *J. Biomech. Eng.* **102**, 42–49 (1980).
47. F. Hirschhaeuser *et al.*, Multicellular tumor spheroids: An underestimated tool is catching up again. *J. Biotechnol.* **148**, 3–15 (2010).
48. X. Wang, H. A. Stone, R. Golestanian, Shape of the growing front of biofilms. *New J. Phys.* **19**, 125007 (2017).
49. M. Martin, T. Risler, Viscocapillary instability in cellular spheroids. *New J. Phys.* **23**, 033032 (2021).
50. D. Hanahan, R. A. Weinberg, Hallmarks of cancer: The next generation. *Cell* **144**, 646–674 (2011).
51. G. Forgacs, R. A. Foty, Y. Shafir, M. S. Steinberg, Viscoelastic properties of living embryonic tissues: A quantitative study. *Biophys. J.* **74**, 2227–2234 (1998).
52. P. A. Netti, D. A. Berk, M. A. Swartz, A. J. Grodzinsky, R. K. Jain, Role of extracellular matrix assembly in interstitial transport in solid tumors. *Can. Res.* **60**, 2497 (2000).
53. R. A. Brace, A. C. Guyton, Interaction of transcapillary Starling forces in the isolated dog forelimb. *Am. J. Physiol.* **233**, H136–H140 (1977).